

# Progress report on an IATTC yellowfin tuna life history project

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# TOPICS

- **Introduction**
- **Objectives**
- **Experimental design**
- **Methodology**
- **Progress and Proposed Timeline**

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- Age and growth of yellowfin in the EPO was investigated, based on otolith samples collected about 40 years ago, with the resulting growth model being used in the current assessment
- Reproductive biology of yellowfin was investigated in the EPO, based on gonad samples collected about 30 years ago, from which maturity and fecundity estimates are being used in the current assessment



# OBJECTIVES

- Obtain current estimates of the age, growth, maturity, and fecundity of yellowfin from throughout the range of the purse-seine fishery in the EPO for utilizing in length-based age-structured stock assessment models

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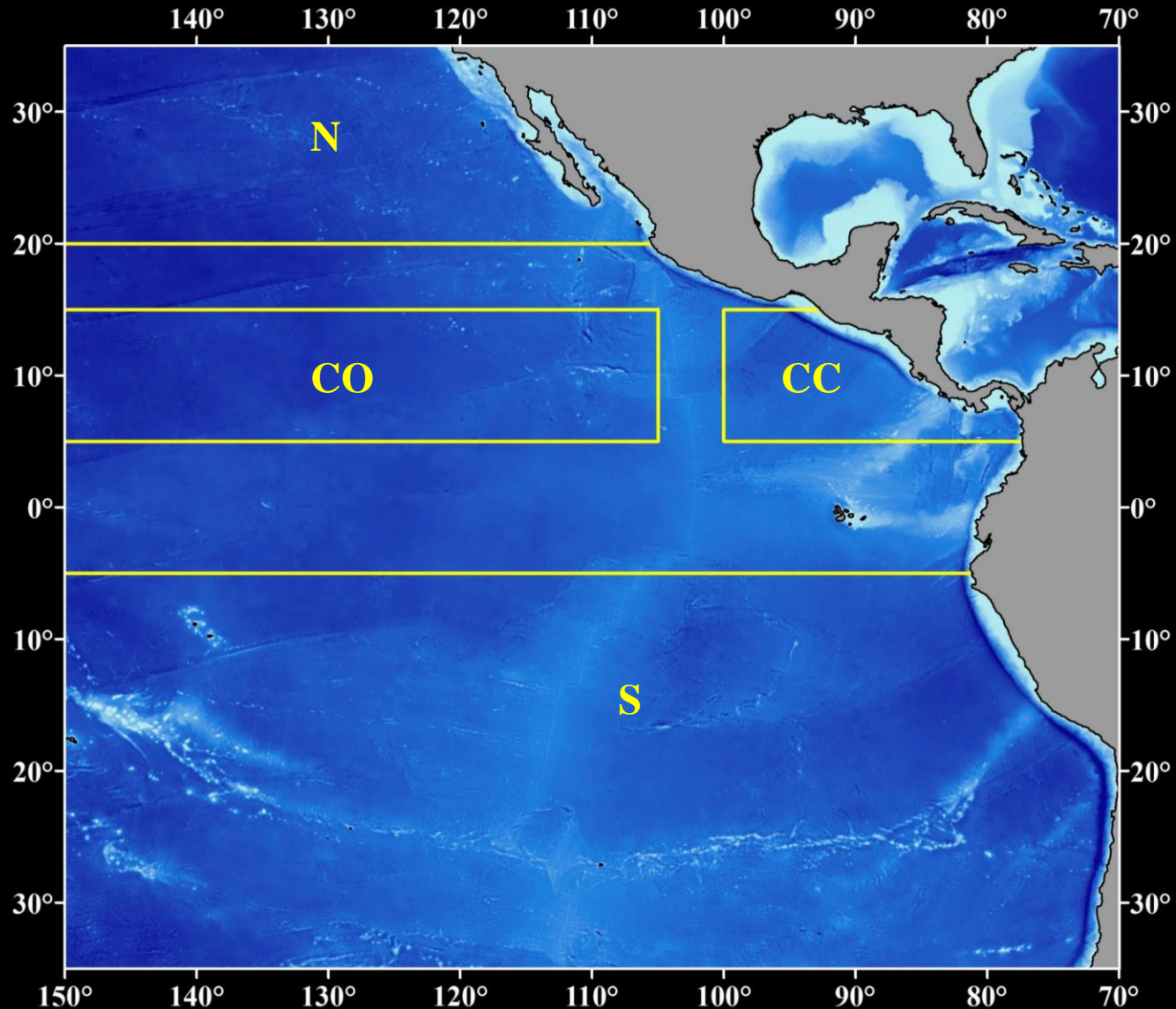
- Obtain current estimates of the age, growth, maturity, and fecundity of yellowfin from throughout the range of the purse-seine fishery in the EPO for utilizing in length-based age-structured stock assessment models
- Evaluate the temporal and spatial variability in these life history characteristics, and their application in spatially explicit yellowfin stock assessment models

# EXPERIMENTAL DESIGN

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4 Sampling areas: North of 20° N, Between 15° N and 5° N from the coast to 100° W, Between 15° N and 5° west of 105° W, and South of 5° S



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- Fish from which ovaries were sampled were tagged, for locating during the unloading process, for collections of heads and extraction of otoliths by IATTC field office staff. Fish lengths were re-measured and their weights recorded at that time.

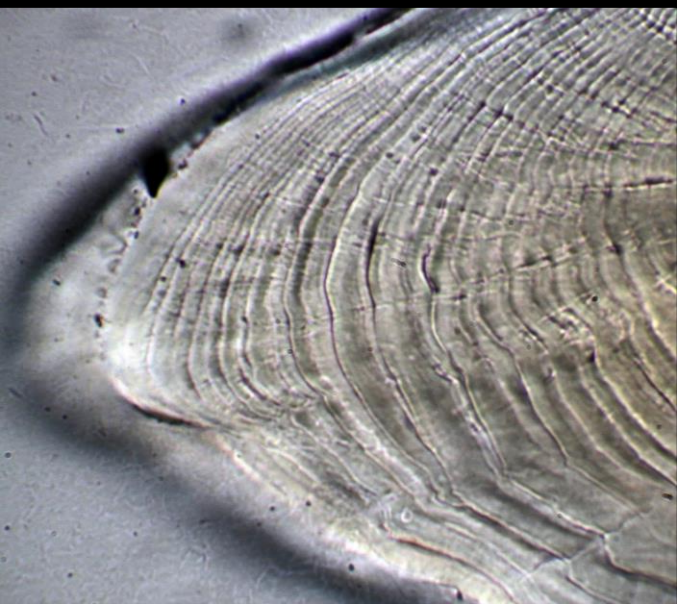
# METHODOLOGY

- Processing of otoliths and ovarian tissues will be undertaken at the IATTC La Jolla Fish Ecology Laboratory

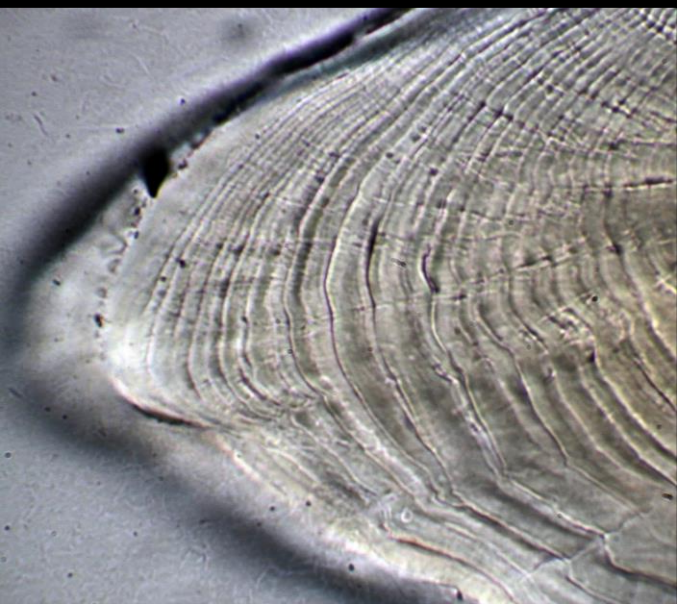
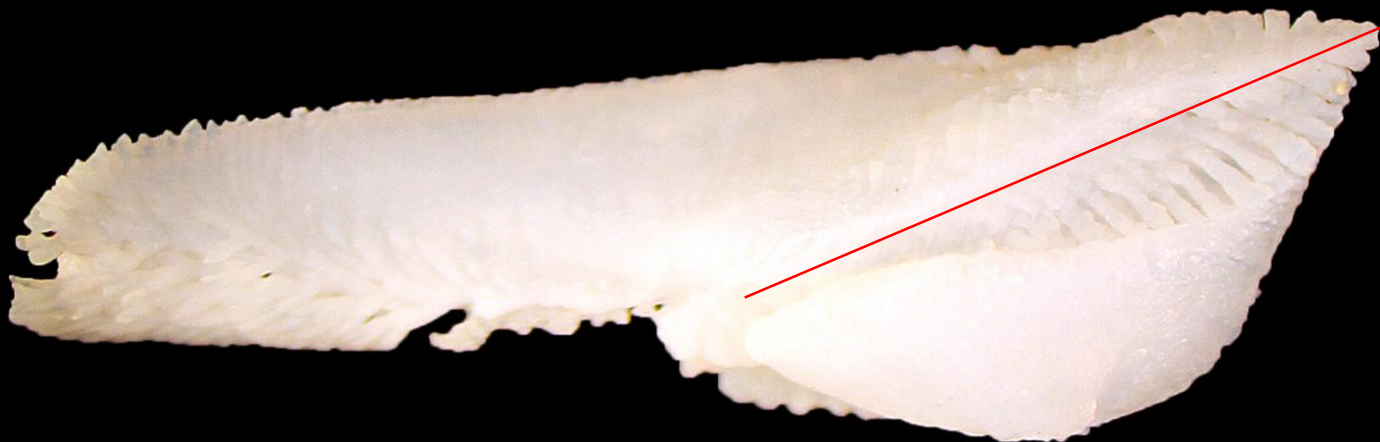
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- Otolith preparation consists of embedding sagittal otoliths in epoxy resin, and cutting frontal sections along the primordium to post-rostral axis, using a low speed saw. Sections are then mounted on microscopic slides, polished to approximately 5 microns in thickness, and then etched with a dilute acid solution to obtain highly visible increments along the optimal counting path

# Yellowfin tuna right sagittal otolith



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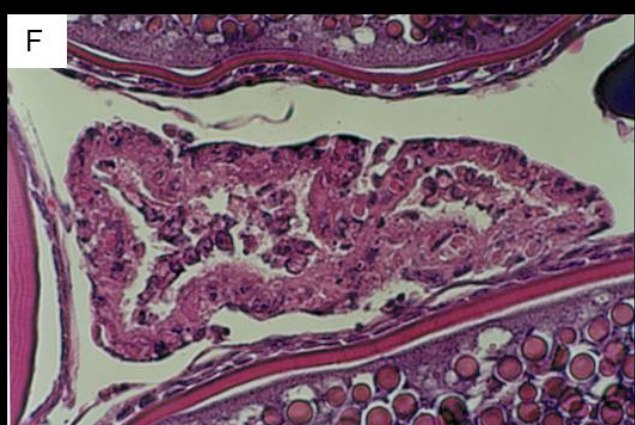
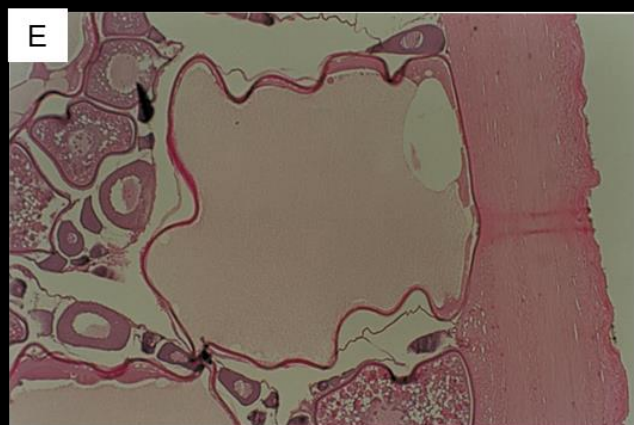
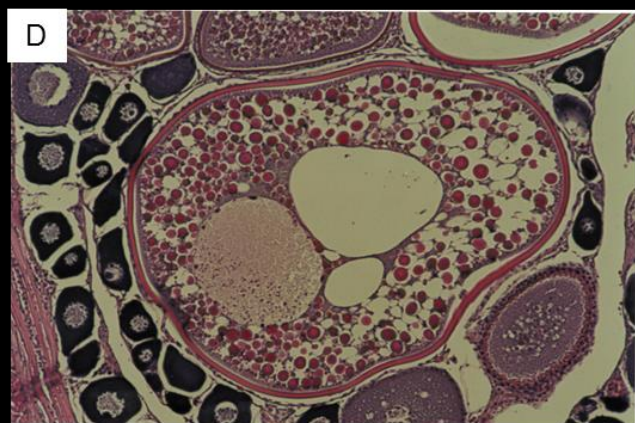
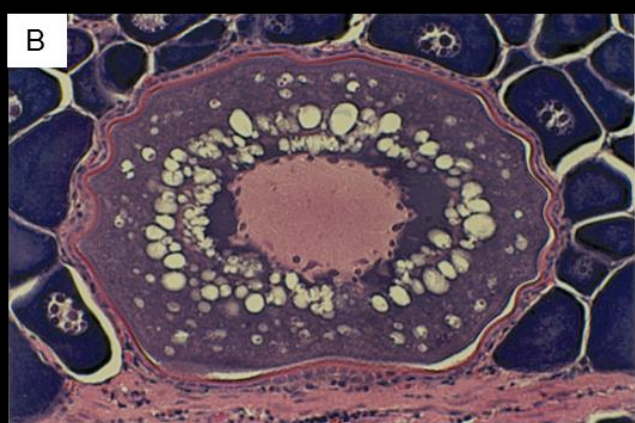
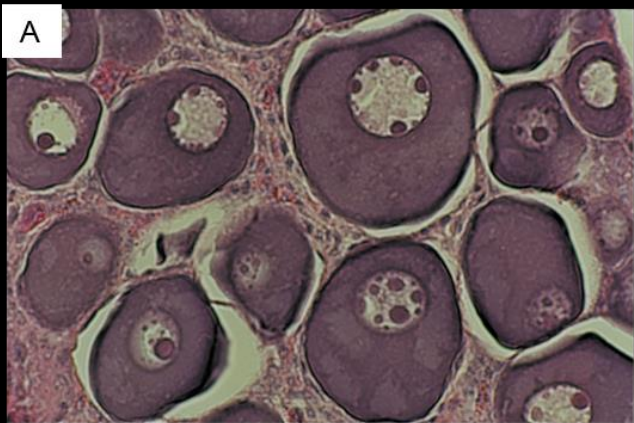
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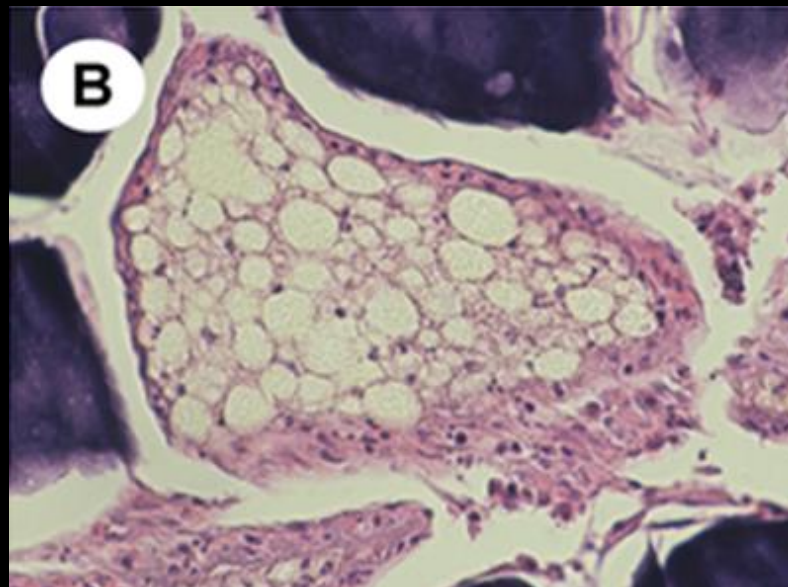
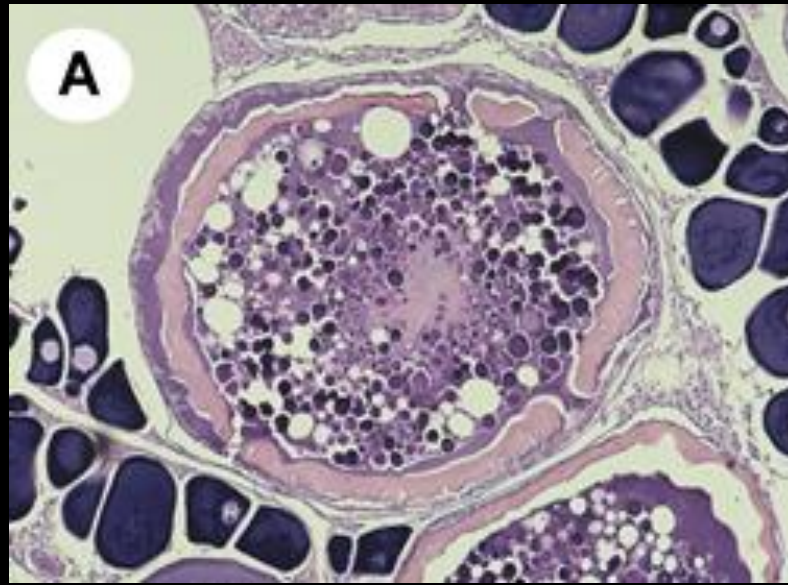
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- For each microscopic slide the histological classification system of Schaefer (1998) will be employed to identify the oocytes in the most-developed mode, absence or presence and age of postovulatory follicles, atretic structures, and additional maturity markers. The histological analyses of ovaries provides accurate evaluations of the maturity and reproductive activity stages for individual females

**Developmental stages and oogenic cells observed in yellowfin ovaries. (A) Unyoked oocyte. (B) Early yoked oocyte. (C) Advanced yoked oocyte. (D) Migratory-nucleus-stage oocyte. (E) Hydrated oocyte. (F) Postovulatory follicle less than 12 h after ovulation**



## Alpha (A) and beta (B) stage atresia in yolked oocytes



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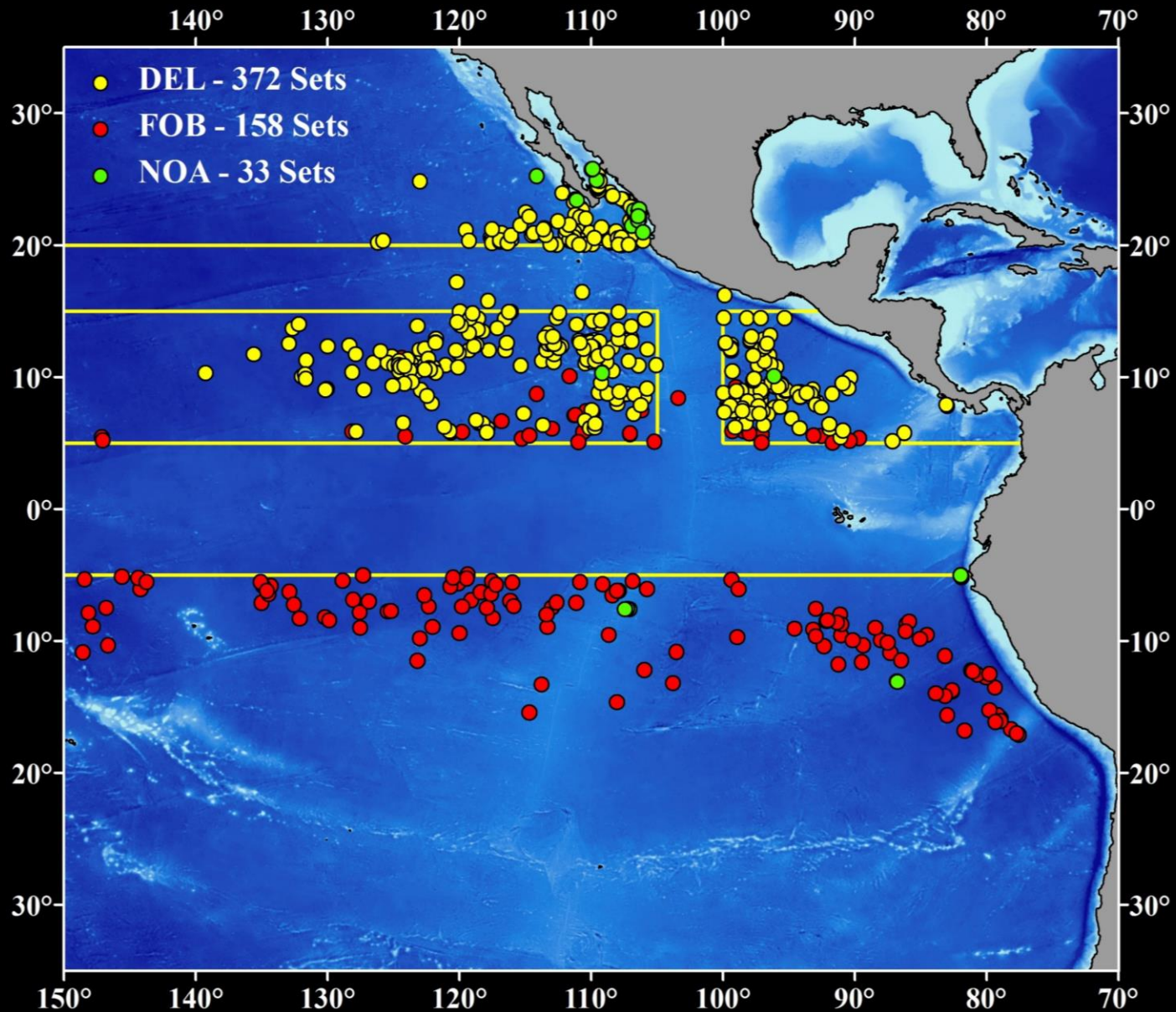
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- Statistical comparisons of the growth, maturity, and fecundity functions obtained from the four spatial strata will be conducted, along with statistical comparisons of those functions derived in the earlier studies

# PROGRESS AND PROPOSED TIMELINE

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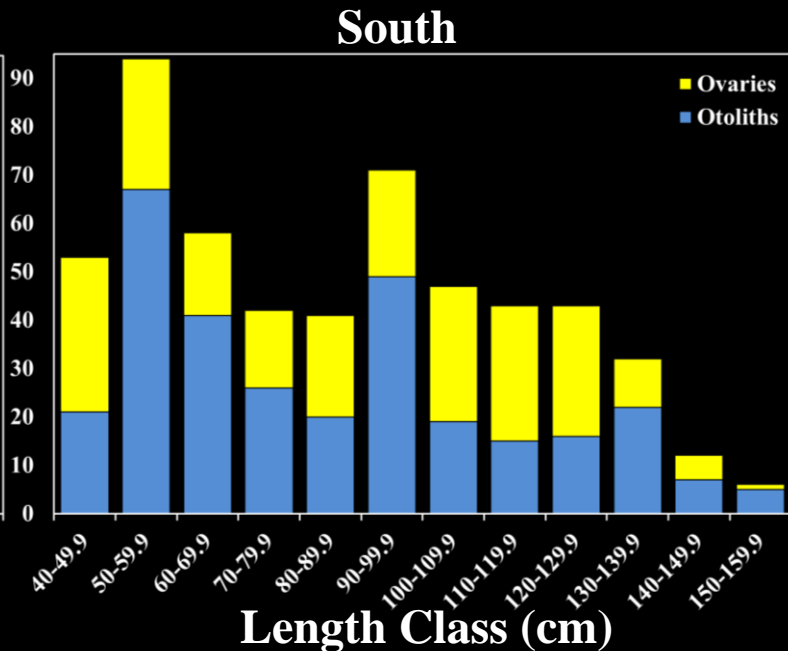
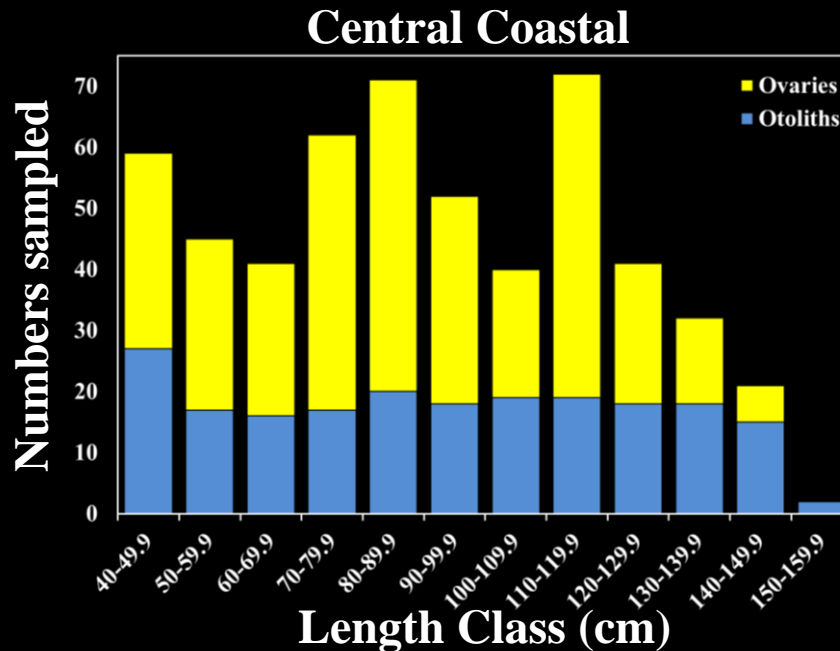
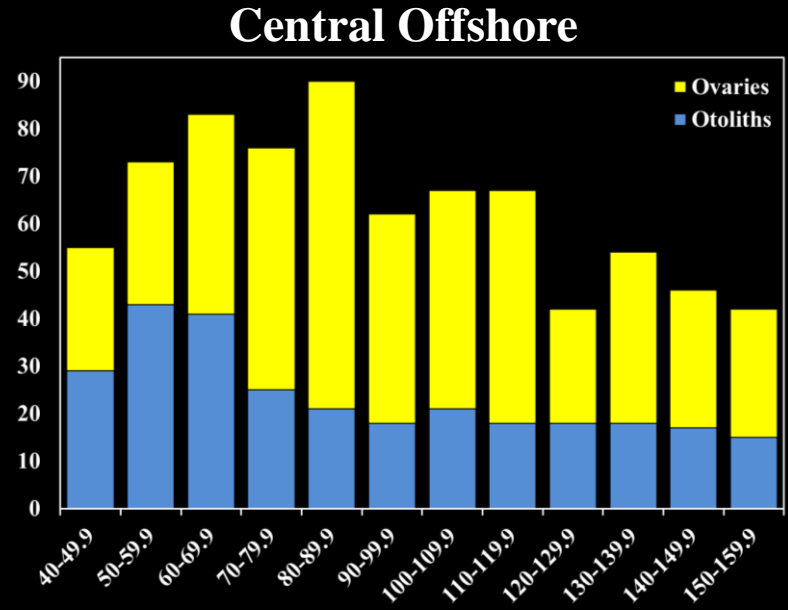
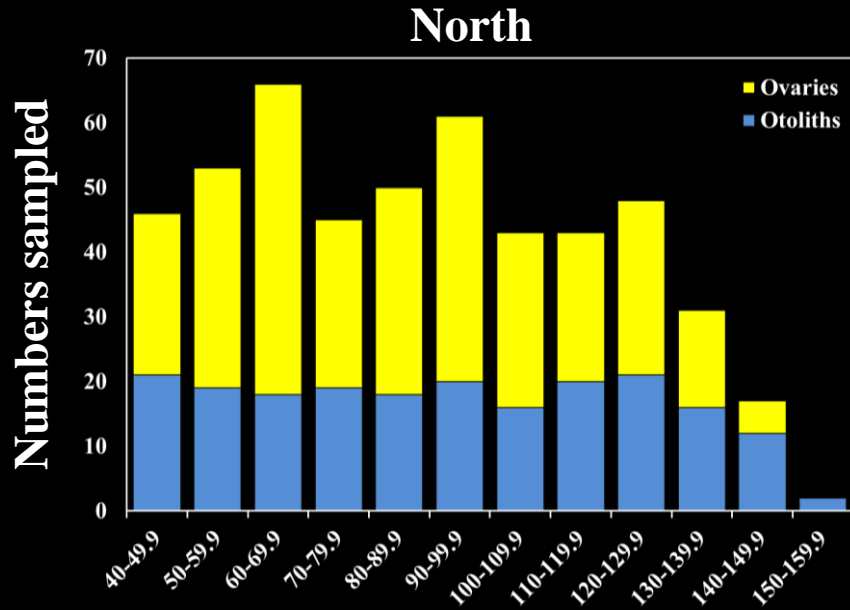
# Position of 563 sets from which ovaries and otoliths were collected



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# Numbers of ovaries and otoliths sampled within the 4 spatial strata



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- Will present an update and results at IATTC SAC9